## 1855242

# Synovial Expression Levels of PD-1, the Target of Rosnilimab, Correlate with Disease Activity and Persist Across Disease Stages and Lines of Therapy in Rheumatoid Arthritis

# ABSTRACT

**Background:** Despite multiple approved therapies in rheumatoid arthritis (RA), many patients do not achieve clinically meaningful responses, emphasizing the need for novel therapeutics with improved efficacy. Rosnilimab is an IgG1 monoclonal antibody that reduces T cell activation through depletion of PD-1 high T cells and agonism of the PD-1 inhibitory receptor. In RA synovium, approximately 80% of T cells are PD-1 positive with an enrichment of CXCL13 secreting PD-1 high T peripheral helper (Tph) cells, which drive B cell differentiation to autoantibody producing plasma cells. To define potential target populations to rosnilimab therapy, we explored synovial PD-1-associated genes and pathways in three RA cohorts synchronized for disease stage and prior treatment exposure: (1) treatment-naive early RA (PEAC); (2) csDMARD-inadequate responders (IRs) (STRAP) and anti-TNF-IRs (R4RA). **Methods:** Transcript expression levels were quantified and normalized using edgeR sea from the above cohorts. Pearson's correlation was used to test associations between gene expression and clinical and imaging features. PD-1 associated pathways were analyzed using Gene Set Variation Analysis (GSVA) on two gene sets: a nivolumab-induced gene set (Choueiri et al 2016) and a rosnilimab-induced gene set generated using RNA-seq data from purified T cells co-cultured with monocyte-derived DCs and rosnilimab **Results:** In treatment naïve patients, synovial PD-1 transcript levels were positively correlated with clinical features at levels, anti-CCP titers, acute phase reactants (CRP p=0.01 and ESR p<0.001), disease activity scores (DAS28-CRP p<0.01 and DAS28-ESR p<0.01), ultrasound synovial thickness (p<0.001), and total Sharp van der Heijde radiographic scores (p<0.001), with positive correlations also observed for CXCL13 transcript levels. Similar trends were observed in DMARD-IR and TNF-IR cohorts, particularly in relation to disease compared to the treatment naïve cohort, PD-1 transcript levels were also expressed in DMARD-IR and TNF-IR cohorts. Despite strong expression of synovial PD-1, anti-nivolumab (PD-1 antagonist) and rosnilimab (PD-1 agonist) gene signatures were not elevated suggesting that insufficient agonism may be contributing to active disease. T cell activation markers and CXCL13 expression levels closely correlated with CD3 expression across disease stages, indicating synovial T cells continue to exhibit an activated, Tph-like phenotype. **Conclusion:** Synovial PD-1 and CXCL13 expression levels correlate with clinical markers of disease and persist across naïve, DMARD-IR, and TNF-IR patients with RA. This supports an important role for PD-1 in disease pathogenesis and the biologic rationale for using rosnilimab following different lines of therapy. This will be tested in the RENOIR study, an ongoing Phase 2 trial of rosnilimab in moderate/severe RA including biologic naïve and experienced patients (NCT06041269).

# **BACKGROUND & OBJECTIVE**

- PD-1 Pathway is Dysregulated in Rheumatoid Arthritis (RA)
   Programmed cell death protein 1 (PD-1) is a coinhibitory receptor that reduces the activation status of T cells when engaged with its ligand PD-L1 <sup>1-3</sup>
- There is over a two-fold increase in PD-1+ T cells in the periphery of RA patients compared to healthy controls<sup>4</sup>
- 80% of T cells in RA synovium are PD-1+<sup>5</sup>

## Rosnilimab (PD-1 modulator, IgG1)

- Mechanism of action and proposed impact on PD-1+ T cells (Fig. 1)
  - Depletion of PD-1<sup>high</sup> Teff, Tfh, and Tph cells and inhibition of remaining PD-1+ T cells resulting in:
  - Reduced pathogenic T cell migration, proliferation, and inflammatory cytokine secretion (e.g.  $IFN\gamma$ )
  - Reduced Tfh and Tph-derived cytokines (IL21 and CXCL13) preventing subsequent plasmablast and plasma cell generation and autoantibody levels
- Modulation through PD-1 may restore immune homeostasis in numerous autoimmune and inflammatory indications, including RA

**Objective:** Assess the therapeutic potential of PD-1 modulation using synovial tissue transcriptomic data from RA patients across different lines of therapy, including treatment naïve as well as inadequate responders to csDMARD and anti-TNF treatments

## METHODS

## **Evaluation of PD-1 expression in RA patient synovial T cells**

- PD-1 levels were evaluated across the single-cell synovial cell atlas from the Accelerating Medicines Partnership (AMP) Phase II Rheumatoid Arthritis study that includes data from 70 patients with various disease severities and treatment histories<sup>6</sup>
- T cell subsets were clustered into 24 subtypes of interest, annotated by the AMP consortium, and assessed for PD-1 expression via single-cell RNA-seq

### Characterization of treatment naïve early RA patient synovial tissue

- The Pathobiology of Early Arthritis Cohort (PEAC) includes RNA-sequencing data from synovial tissue biopsied from 90 early RA patients naïve to therapy<sup>7</sup>
- The Gene View module on the QMUL PEAC RNA-seq Data shiny website was utilized to evaluate PD-1 and CXCL13 expression correlation with clinical phenotypes

## Analysis of PD-1 levels and T cell activation status in RA patient synovium across different lines of therapy

- RNA-seq data across RA patient synovium (n=20)/cohort from three different study cohorts; PEAC, STRAP (csDMARD inadequate responders), and R4RA (TNF inadequate responders) were analyzed for CD3 expression and compared to PD-1 expression and markers of Tph and T cell activation
- Transcriptomic data of osteoarthritis (OA) patient synovial tissue (n=6) from GEO accession GSE254682 was analyzed and served as a control group

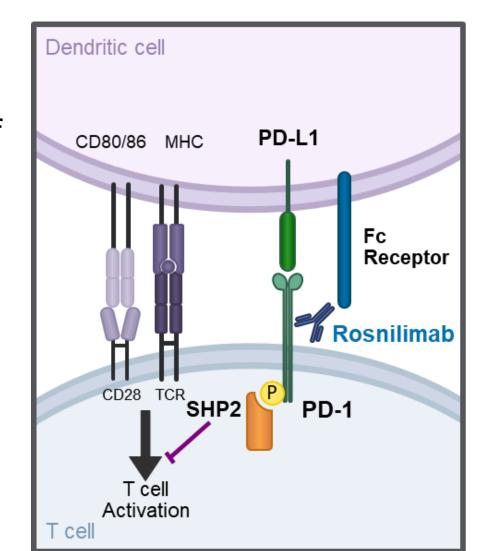


Figure 1. Proposed MoA of rosnilimab

Yangsu Ren<sup>1</sup>, Catherine Aversa<sup>1</sup>, Myles Lewis<sup>2</sup>, Cankut Cubuk<sup>2</sup>, Felice Rivellese<sup>2</sup>, Liliane Fossati-Jimack<sup>2</sup>, Pejman Soroosh<sup>1</sup>, Amy M. Archer<sup>1</sup>, Martin Dahl<sup>1</sup>, Paul Lizzul<sup>1</sup>, Cailin Sibley<sup>1</sup>, and Costantino Pitzalis<sup>2</sup> <sup>1</sup>AnaptysBio, San Diego, United States

<sup>2</sup>Centre for Experimental Medicine and Rheumatology, Queen Mary University of London, London, UK

## PD-1 was Highly Expressed on Tph/Tfh, Proliferating, and Memory T Cells in RA Patient Synovium

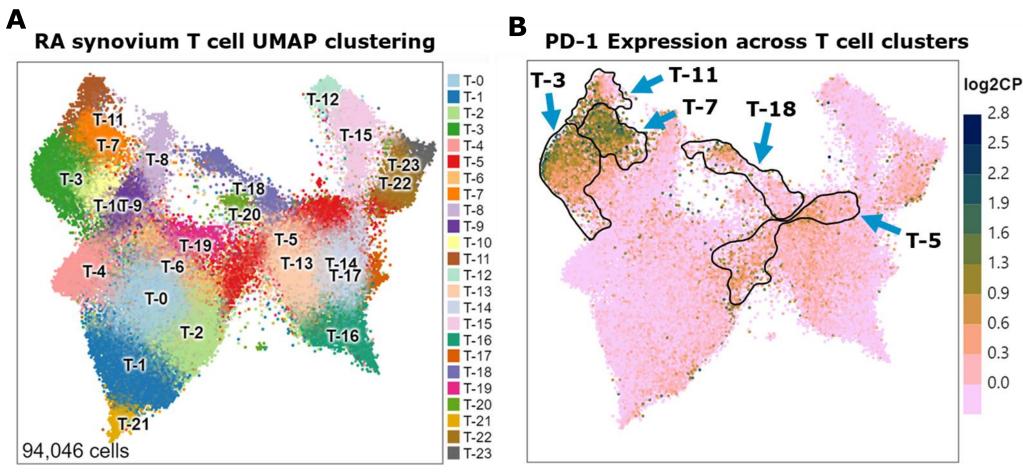


Figure 2. Uniform manifold approximation and projection (UMAP) clusters of T cells from RA patient synovium with arrows identifying Tph and Tfh/Tph cells, T-7 and T-3, respectively (A) and feature plot of PD-1 expression across T cell subtypes (B)

## **PD-1 and CXCL13 Expression in RA Synovium was Significantly Correlated** with RA Disease Activity

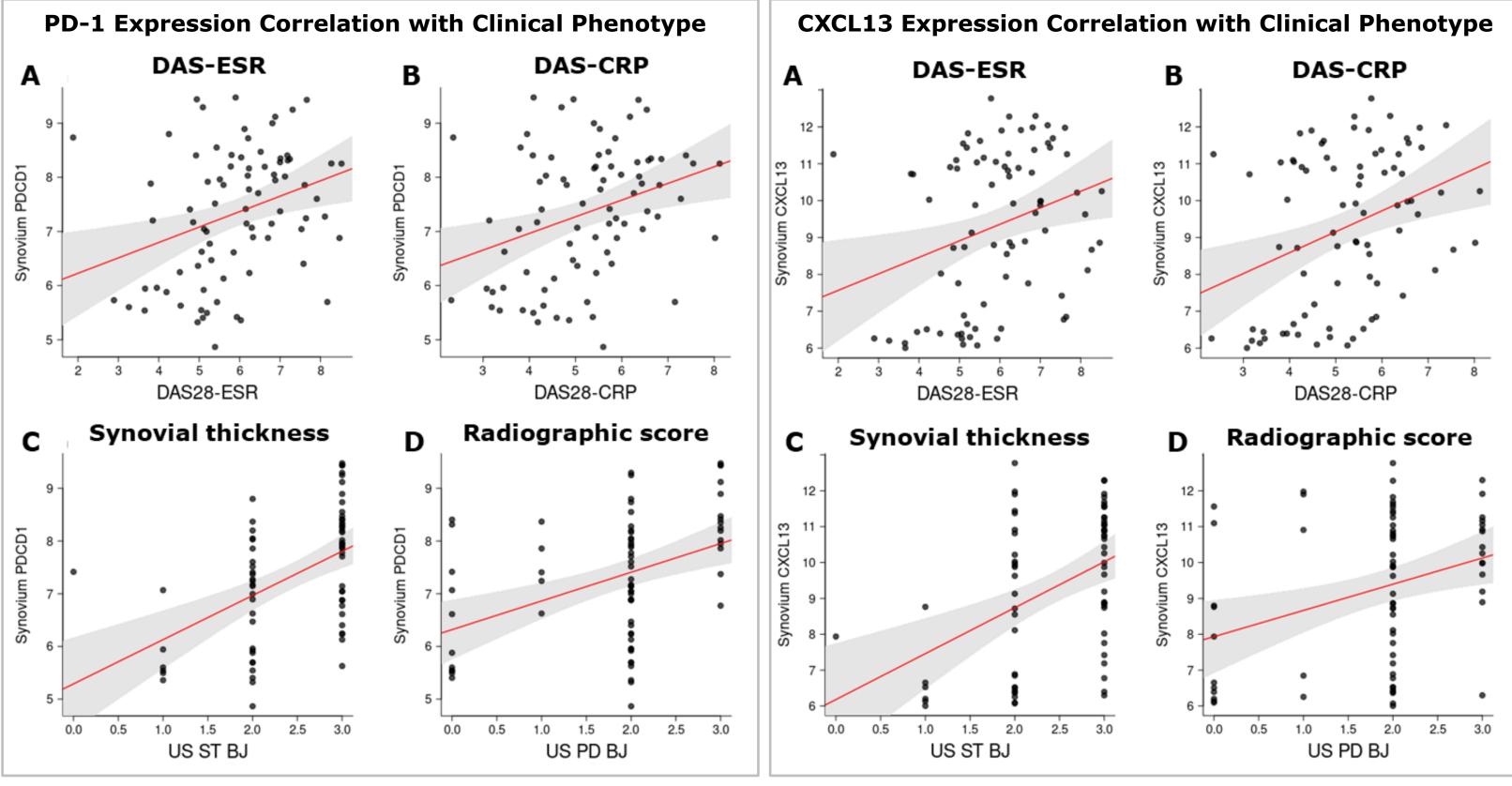


Figure 3. Correlation of PD-1 with RA patient disease activity scores DAS82-ESR (A) and DAS28-CRP (B), ultrasound synovial thickness (C) and total Sharp van der Heijde radiographic score (D)

From treatment naïve early RA patient-derived synovial biopsy bulk RNA-seq (PEAC cohort): • Both PD-1 and CXCL13 transcript levels showed significant positive correlations with disease activity scores (Fig. 3A & 4A) and DAS28-ESR (Fig. 3B & 4B)

- PD-1 and CXCL13 also both significantly correlated with ultrasound synovial thickness (US ST BJ Fig. 3C & 4C), and total Sharp van der Heijde radiographic scores (US PD BJ Fig. **3D & 4D**)

# RESULTS

- AMP RA phase II patient-derived synovial T cells were clustered into 24 unique T cell subtypes via UMAP analyses (**Fig. 2A**)
- CD4+ Tph (T-7) and Tfh/Tph (T-3) cells demonstrated the highest PD-1 expression compared to other T cell subsets (**Fig. 2B**)
- Notably, high PD-1 expression was also observed across proliferating T cells (T-18) and several memory T cell subtypes, including CD4+ CD146+ memory (T-11) and CD4+ GZMK+ memory (T-5) T cells

Figure 4. Correlation of CXCL13 with RA patient disease activity scores DAS82-ESR (A) and DAS28-CRO (B), a ultrasound synovial thickness (C) and total Sharp van der Heijde radiographic score (D)

### **Correlation P-values**

	DAS-ESR	DAS-CRP	Synovial Thickness	Radiographic score
PD-1	2.2x10 <sup>-3</sup>	4.8x10 <sup>-3</sup>	4.9x10 <sup>-6</sup>	7.8x10 <sup>-5</sup>
CXCL13	3.0x10 <sup>-3</sup>	1.9x10 <sup>-3</sup>	1.9x10 <sup>-4</sup>	1.2x10 <sup>-2</sup>

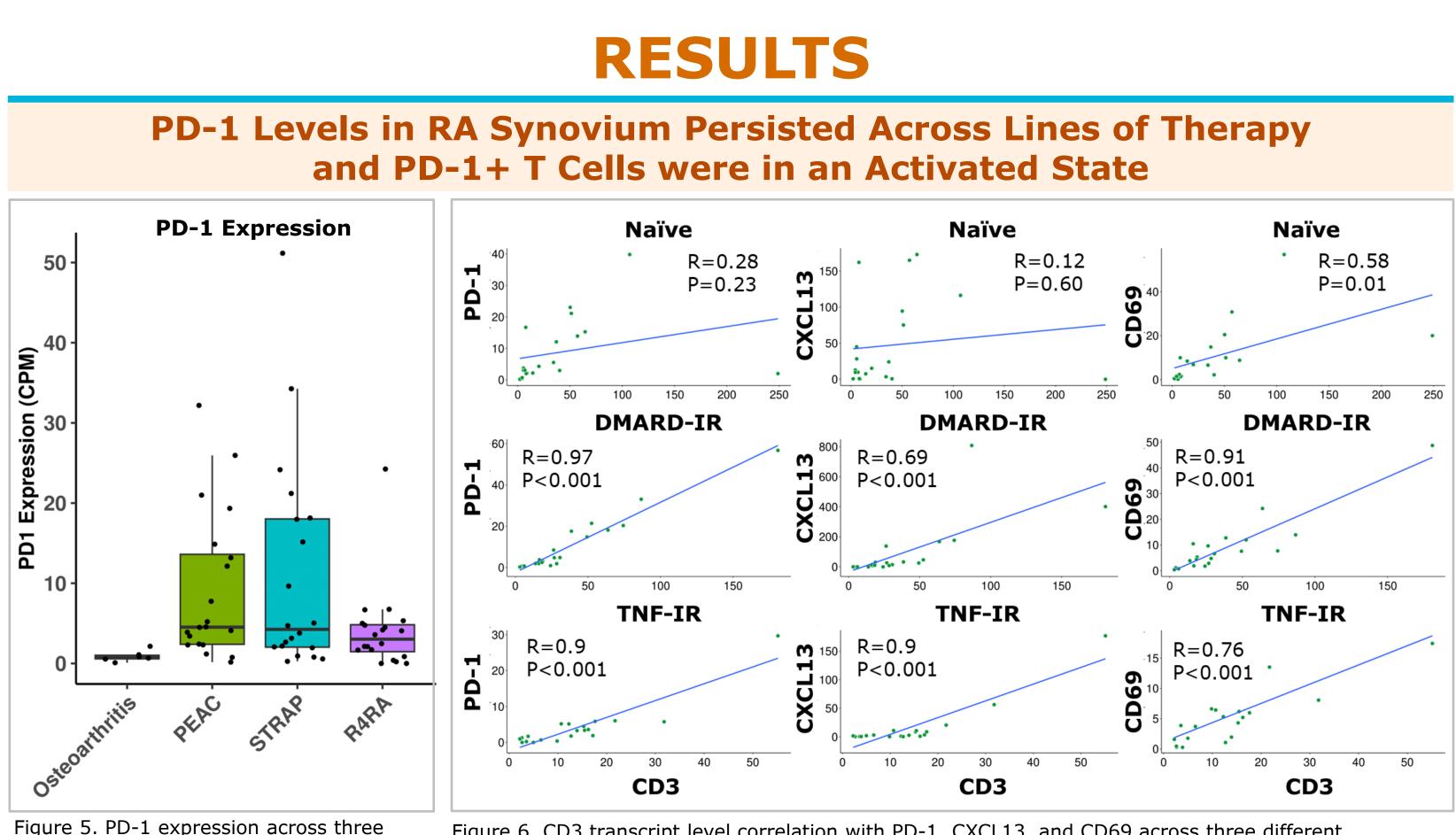


Figure 6. CD3 transcript level correlation with PD-1, CXCL13, and CD69 across three different Figure 5. PD-1 expression across three different treatment cohorts compared to OA controls

- Tph-like phenotype (Fig. 6)

## and joint inflammation

- of therapy
- of rosnilimab (NCT06041269)

## 1. This research was funded by Anaptys

## 1. Okazaki T, Honjo T. Trends Immunol 2006;27:195–201.

- 2. Chen et al. Front Immunol 2023; 16:14:1163633.
- 3. Gao M, et al. *Cancer Letters* 2024; 588: 216726
- 4. Chen et al. *Clin Transl Imm* 2024;13(10).

• PD-1 transcript levels show upregulation in RA patients compared to OA controls, and maintained expression levels in patients post-treatment with csDMARD and anti-TNF therapies (Fig. 5)

• T cell activation markers (PD-1 and CD69) and CXCL13 expression levels closely correlated with CD3 expression across disease stages, indicating synovial T cells continue to exhibit an activated,

# CONCLUSION

Synovial PD-1 and CXCL13 expression levels correlated with clinical markers of disease activity

• PD-1+ T cells in RA synovium were observed to be preferentially in an activated state and persisted across naïve, DMARD-IR, and TNF-IR patients

• These data support the importance of PD-1 in RA disease pathogenesis and the biological rationale for developing rosnilimab as a potential treatment option following other lines

• RENOIR, an ongoing Phase 2 trial of rosnilimab in moderate-to-severe RA, includes biologic naïve and experienced patients that will provide additional insight on the therapeutic potential

# ACKNOWLEDGEMENTS

2. Cynthia Alexander of Anaptys provided medical writing support

3. Disclosures: ML, CC, FR, LFJ, and CP are consultants and received research grants from Anaptys. For ML, CC, FR, LFJ, and CP, additional disclosures available on request. All other authors are employees and stockholders of Anaptys

# REFERENCES

5. Aletaha and Smolen, JAMA 2018;320(13):1360.
6. Zhang et al. Nature 2023;623(7987):616-624.
7. Lewis et al. Cell Reports 2019;28(9): 2455-2470.e5.

